# Enhancing Leaf Flavonoid Production in Indian Camphorweed (*Pluchea indica* Less.) through the Provision of Chicken Manure

# Yulia Indriani<sup>A</sup>, Sandra Arifin Aziz\*BC, Maya Melati<sup>BC</sup>

- <sup>A</sup>Postgraduate School, Agronomy and Horticulture, Bogor Agricultural University, Bogor, Indonesia, 16680
- <sup>B</sup> Department of Agronomy and Horticulture, Faculty of Agriculture, Bogor Agricultural University, Bogor, Indonesia, 16680
- <sup>c</sup> Biopharmaca Research Center, Bogor Agricultural University, Bogor, Indonesia
- \*Corresponding author; sandraaziz@apps.ipb.ac.id

## **Abstract**

Indian camphorweed (Pluchea indica Less.) leaves exhibit antioxidant, antibacterial, anti-inflammatory, and antimicrobial activities, indicating significant potential for the pharmaceutical industry. This research aimed to determine the optimal rates of chicken manure for enhancing leaf and total flavonoid production in Pluchea indica. The study took place at the IPB Experimental Station in Bogor, Indonesia, spanning from July to October 2023. A completely randomized block design with a single factor (chicken manure doses) was employed: 0, 2.5, 5, or 7.5 kg per plant. Each treatment comprised three replications, each consisting of 20 plants. The results revealed that the application of chicken manure at 2.5 kg per plant led to significantly taller plants, more leaves, and tertiary branches compared to the control group. Specifically, the 2.5 kg dosage significantly increased the nitrogen content of the 7th leaf and the potassium content of the 3rd leaf. Meanwhile, the application of 5 kg of chicken manure per plant significantly boosted phosphorus content in the 3<sup>rd</sup> and 7<sup>th</sup> leaves and potassium content in the 5th and 7th leaves. However, no significant differences were observed in total flavonoid and antioxidant activity across all leaf positions with chicken manure application. Plants receiving 5 kg of manure demonstrated the highest fresh leaf weight (81.64 g) and dry weight (38.27 g), which were significantly greater than those receiving 2.5 kg per plant or no manure. Despite these variations, flavonoid production per plant did not show a significant difference with manure application.

Keywords: anti-bacterial, anti-inflammatory, antioxidant, functional food, organic

## Introduction

Indian camphorweed, Indian fleabane, or Indian pluchea (Pluchea indica Less.) is an indigenous Asteraceae species utilized as a functional food, with a limited number of individuals incorporating its leaves as a food supplement. Recognized as a shrub capable of attaining a height of 2 meters, Pluchea indica is indigenous to a vast region encompassing much of Asia, India, and northern Australia (Parker, 2012). The leaf extract of Pluchea indica is rich in flavonoids (Yuliani et al., 2015; Tinrat, 2021), alkaloids (Mutrikah et al., 2018), saponins (Tinrat, 2021), and tannins (Lestari et al., 2020; Tinrat, 2021). Notable compounds include 1,3,4,5-tetra-O-caffeoylquinic acid and 3,4,5-tri-O-caffeoyl quinic acid (Ohtsuki et al., 2008). Within the flavonoid group of P. indica leaves, quercetin and kaempferol are the primary constituents (Suriyaphan, 2014), with the flavonoid concentration containing 81% quercetin (Andarwulan et al., 2010).

The diverse contents of *Pluchea indica* leaves include antibacterial properties (Srimoon and Ngiewthaisong, 2015; Lestari et al., 2020), anti-inflammatory effects (Pranata et al., 2021), as well as antimicrobial and antioxidant activities (Tinrat, 2021). The leaf extract of Pluchea indica finds applications in the formulation of products such as lotion bars (Tinrat, 2021), facial soaps (Komala et al., 2020), and anti-acne serums (Komala et al., 2021).

Best practices for the cultivation of *Pluchea indica* are not well-established, despite its extensive use as a medicinal plant with potential applications in the cosmetic and pharmaceutical industries. Minister of Agriculture Regulation Number 64/Permentan/OT.140/5/2013 recommends the cultivation of medicinal plants using organic fertilizer (Kementan, 2013). One such organic fertilizer is chicken manure,

typically containing 1.64% total N, 5.14% total P, 1.6% K2O, and 19.12% C-organic (Betty, 2018). Application of chicken manure to ultisol has been shown to improve soil chemical properties, including soil pH, C-organic, total N, C/N ratio, P-available, and CEC (Walida et al., 2020). Chicken manure provides nutrients that enhance the physical and chemical properties of the soil, supporting the cultivation of plants.

The nitrogen content in chicken manure serves as a substrate for the enzyme phenylalanine ammonia-lyase (PAL), a key enzyme in the flavonoid biosynthesis pathway (Roy et al., 2022). Nitrogen is essential for regulating flavonoid biosynthesis by allocating carbohydrates for both primary and secondary metabolism (Deng et al., 2019). Phosphorus in chicken manure acts as a substrate for phosphoenolpyruvate (PEP) and D-erythrose 4-phosphate (E4P) compounds, forming 3-Deoxy-D-arabino-heptulosonic acid 7-phosphate (DAHP) precursors in the Shikimate pathway, which participates in flavonoid biosynthesis (Tariq et al., 2023). Potassium serves as an activator for crucial enzymes involved in protein synthesis, sugar transport, and photosynthesis (Marschner, 2012).

Several studies have reported positive effects of chicken manure in increasing nutrient and flavonoid content in plants. For instance, chicken manure application to Moringa increased leaf production and total flavonoids (Rasmani, 2021). Research by Karimuna (2015) demonstrated that the highest levels of total flavonoids in orange jessamine leaves were obtained with chicken manure doses of 5 and 7.5 kg per plant. The optimal dose of chicken manure to increase the production of Vernonia amigdala leaf flavonoids was found to be 7.5 kg per plant (Betty, 2018). Previous research on orange jessamine plants indicated that while applying chicken manure increased the fresh and dry weight of leaves, it reduced the flavonoid concentration (Karimuna et al., 2015). Hence, this study aims to investigate if a similar effect might occur in Pluchea indica. Consequently, the research aims to determine the optimal application dose of chicken manure to increase leaf production and total flavonoids in Pluchea indica.

## **Material and Methods**

The research was conducted from July to October 2023 at the Cikarawang experimental field, IPB, Bogor, West Java, Indonesia. Leaf tissue analysis was carried out at the AGH-IPB Testing Laboratory. The leaves were harvested at 8 WAP (week after planting). Fresh and dry leaf weights were measured.

The total flavonoid analysis was carried out at the post-harvest laboratory, Department of Agronomy and Horticulture, IPB. Antioxidant activity analysis was carried out at the Biochemistry Laboratory, IPB.

The research used 2-month-old cuttings as planting materials. Rooted cuttings were planted in the field with a planting space of 1 m x 1 m in 4 m x 5 m plots. Each planting hole was applied with chicken manure according to the treatments.

#### Treatment

This research used a completely randomized block design with one factor/treatment, the rate of chicken manure was 0, 2.5, 5, and 7.5 kg per plant and repeated three times, totalling 12 experimental units; each the experimental unit consists of 20 plants.

## Fresh and Dry Leaf Weight

Plants were harvested when having a minimum height of 45 cm from the ground surface and then pruned at 30 cm height. All leaves were separated from the branches to measure the leaf fresh and dry weights. The leaf positions of the 3<sup>rd</sup>, 5<sup>th</sup>, and 7<sup>th</sup> from the shoot of each pruned branch were separated for further analysis. The harvested leaves were air-dried and then placed in the oven at 70° C for 2 x 24 hours.

# Antioxidant Activity Analysis

Analysis of antioxidant activity used dry powder leaves from the 3<sup>rd</sup>, 5<sup>th</sup>, and 7<sup>th</sup> leaf positions using 1,1-diphenyl-2-picrylhydrazyl (DPPH) as a free radical scavenging assay (Fukumoto and Mazza, 2000; Karimuna et al., 2015). Leaves were dried in an oven at 60° for 3 x 24 hours prior to analysis. The 0.1-q powder was extracted in 1 ml of MeOH in a microtube and then heated for 60 minutes at 60° C. Then the mixture was centrifuged for 5 minutes at 5° C at 12000 rpm and the supernatant was separated. DPPH was dissolved in 80% methanol 0.1 mM. The standard solution uses ascorbic acid in various concentrations (0-100 µg.ml-1). The sample was placed in a 96well microplate reader with 22 µl and added 200 µl of DPPH solution and triplicate. The blank used 200 μl of DPPH added with 22 μl of 80% methanol and carried out three times. The mixed solution was kept in a dark place for 20 minutes at 27° C, the solution was vortexed and measured at a wavelength of 517 nm using a nano spectrometer. Antioxidant activity was measured from the percentage of inhibition with the formula:

Percentage of inhibition of DPPH activity =  $(A - B) / A \times 100\%$ 

Where A = control inhibition, and B= sample inhibition.

## Total Flavonoid Analysis

Analysis of total flavonoids used Vongsak et al. (2013) with a modification from Chang et al. (2002) using fresh samples of P. indica leaves in positions 3<sup>rd</sup>, 5<sup>th</sup>, and 7<sup>th</sup> on the secondary branch of each replication. Fresh leaves (0.001 g) were crushed and macerated with 2 ml of 70% ethanol. A sample of 0.5 ml was mixed with 0.5 ml of 2% aluminum chloride solution, potassium acetate (0.1 ml, 1 mM). and 2.8 ml of distilled air against the blank sample without aluminum chloride. The standard solution is 10 milligrams of quercetin dissolved in 80% ethanol which was diluted to 0, 2, 5, 25, 50, and 100 μg.ml<sup>-1</sup>. The standard solution was mixed using a vortex, then incubated at room temperature 27° C for 30 minutes. The absorbance of the mixture was measured at 415 nm with a Shimadzu UV-160A spectrophotometer. The standard curve constructed was y = 0.03021 +0.01006, R2 = 0.99977. The results were equivalent to mg of quercetin per gram of fresh sample. The total flavonoid per plant was obtained by multiplying the fresh leaf weight per plant with the total flavonoid concentration of the 7th leaf.

# Leaf Tissue Analysis

Analysis of nitrogen, phosphorus, and potassium concentration was conducted in leaves at positions  $3^{\rm rd}$ ,  $5^{\rm th}$ , and  $7^{\rm th}$  from the apex of each plant. Total N concentration was determined using Kjeldhal method. Phosphorus and potassium concentration were extracted by wet ash method using a mixture of concentrated HNO $_3$  and HCIO $_4$ . Phosphorus concentration was determined using a spectrophotometer, whereas potassium using an atomic absorption spectrophotometer (AAS) (Balitttanah, 2005).

#### Plant Growth Measurement and Data Analysis

Growth variables consisted of plant height, leaf number, measured weekly at 2 until 8 weeks after planting (WAP); branch number data was collected at 6 until 8 WAP. Nutrient levels of leaves, total flavonoid, and antioxidant activity were measured at the 3<sup>rd</sup>, 5<sup>th</sup>, and 7<sup>th</sup> leaf positions from the apex. Yield was measured at 8 WAP or when plants reach a minimum height of 45 cm above the ground.

Data were analyzed using the F-test at a 5% significance level, followed by the Duncan Multiple Range Test. If the value of coefficient of variance was more than 30%, data was transformed using the formula  $\sqrt{x+0.5}$ . Statistical analysis used R Studio version 4.3.1 software.

# **Result and Discussion**

Plant Height, Leaf Number, and Branch Number

Providing chicken manure affects the vegetative growth of *P. indica* (Table 1 and Table 2). Plants applied with 2.5 kg of chicken manure showed better growth compared to those without manures, specifically in plant height, leaf number, and number of tertiary branches. Increasing the dose of chicken manure significantly increased plant height (P<0.05) and leaf number (P<0.05) at 6 weeks after planting (WAP), but similar with 5 and 7.5 kg chicken manure per plant. Application of chicken manure also increased the number of tertiary branches (P<0.05) at 8 WAP.

Table 1. Effect of chicken manure application on plant height and leaf number of Pluchea indica

Chicken	Plant height (cm)				Leaf number			
Manure (kg per plant)	2 WAP	4 WAP	6 WAP	8 WAP	2 WAP	4 WAP	6 WAP	8 WAP
0	15.61	29	41.61 b	46.37 b	35.56	214.01	511.73 b	689.46 b
2.5	16.46	34.13	52.28 a	63.93 a	42.33	282.11	852.17 a	1,268.39 a
5.0	16.76	34.98	53.22 a	66.23 a	47.83	279.61	779.28 a	1,361.06 a
7.5	15.05	30.90	48.56 a	74.67 a	31.39	250.67	701.61 a	1,198.67 a
P value	0.2300	0.2020	0.0152	0.0031	0.0634	0.1890	0.0138	0.0055
Sig.	ns	ns	*	**	ns	ns	*	**

Note: Values followed by the different letters in the same column are significantly different in the Duncan test  $\alpha$ =0.05; nsnonsignificant; WAP: week after planting.

Table 2. Effect of chicken manure application on branch number of Pluchea indica.

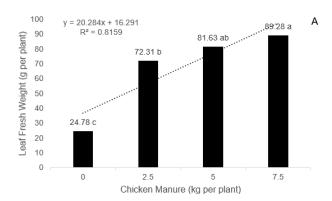
	Branch number						
Chicken Manure (kg per plant)	Prin	Primary		ndary	Tertiary		
(kg per plant)	6 WAP	8 WAP <sup>1)</sup>	6 WAP	8 WAP	6 WAP <sup>1)</sup>	8 WAP	
0	4.60	5.97	27.71	34.91	21.67	29.08 b	
2.5	4.28	6.94	41.11	65.22	35.33	98.47 a	
5.0	4.89	6.17	32.17	66.50	32.54	86.50 a	
7.5	4.56	5.22	25.33	45.11	27.24	109.10 a	
P value	0.936	0.784	0.173	0.173	0.194	0.020	
Sig.	ns	ns	ns	ns	ns	*	

Note: Values followed by the different letters in the same column are significantly different in the Duncan test  $\alpha$ =0.05; nsnonsignificant; <sup>1)</sup> Data was transformed using the formula  $\sqrt{x+0.5}$ ; WAP: week after planting.

## Leaf Fresh and Dry Weight

Chicken manures increased leaf fresh and dry weight of *P. indica* (Figure 1). Plants that received a dose of 5 kg of manure per plant had higher fresh (A) and dry (B) leaf weights compared to those received 2.5 kg per plant or without chicken manure.

per plant with the total flavonoid concentration at the 7<sup>th</sup> leaf position. Flavonoid production per plant also did not have a difference due to the application of chicken manure. In general, the total flavonoid concentration in the 3<sup>rd</sup> leaf position had a greater value than the 5<sup>th</sup> and 7<sup>th</sup> leaf positions as well as the antioxidant activity in the 3<sup>rd</sup> leaf position.



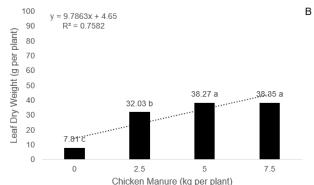


Figure 1. Effect chicken manure application on fresh and dry weight of leaves Pluchea indica

## Leaf Nutrient Concentration

Provision of chicken manure increased the nutrient concentration in leaf tissue. At 2.5 kg per plant N levels of the  $7^{th}$  leaf and K levels of the  $3^{rd}$  leaf increased (Table 3). Application of 5 kg chicken manures per plant significantly increased the P levels of the  $3^{rd}$  and  $7^{th}$  leaves and the K levels of the  $5^{th}$  and  $7^{th}$  leaves (Table 3).

Flavonoid Production, Antioxidant Activity, and Flavonoid Production

The application of chicken manure did not affect the total flavonoid concentration and antioxidant activity at each leaf position (Table 4). There were no differences in total flavonoid levels and antioxidant activity in various leaf positions. Production of total flavonoids per plant was obtained by multiplying the fresh weight

#### **Discussion**

In another Asteraceae species, Vernonia amygdalina, chicken manure increased plant height, branch number, fresh leaf weight, and dry leaf weight (Tjhia et al., 2018). Chicken manure contains 1.64% N, 5.14% total P, 1.6% K2O, and 19.12% C-organic (Betty, 2018). All these nutrients have major roles in stimulating plant vegetative growth. Nitrogen is an essential element for the synthesis of amino acids, proteins, and nucleic acids (Taiz and Zeiger, 2002). Nitrogen increases the amount of chlorophyll thereby encouraging photosynthesis. Nitrogen increases the photosynthesis apparatus so that it can increase the chlorophyll content, the number and activity of carboxylation enzymes, total protein, sugar content, total N, and metabolites related to photosynthesis (Bassi et al., 2018). Phosphorus is essential for energy transfer (Tian et al., 2019), photosynthesis,

Table 3. Effect chicken manure application on concentration N, P, and K leaf of *Pluchea indica*.

Chicken	Nutrient	Leaf position				
manure (kg per plant)	concentration (%)	3 <sup>rd</sup>	5 <sup>th</sup>	7 <sup>th</sup>		
0		3.87	3.92	3.78 b		
2.5	N.I.	4.11	3.97	4.08 a		
5.0	N	4.14	4.06	4.08 a		
7.5		4.26	4.24	4.25 a		
P value		0.1627	0.1720	0.0192		
Sig.		ns	ns	*		
0		0.27 b	0.24	0.22 c		
2.5	D	0.28 b	0.25	0.24 bc		
5.0	Р	0.31 ab	0.29	0.27 ab		
7.5		0.34 a	0.30	0.29 a		
P value		0.0328	0.0522	0.0328		
Sig.		*	ns	*		
0		1.04 b	1.00 c	0.98 c		
2.5	IZ.	1.48 a	1.30 b	1.26 b		
5.0	K	1.62 a	1.71 a	1.48 a		
7.5		1.59 a	1.55 ab	1.54 a		
P value		0.0098	0.0030	0.0023		
Sig.		**	**	**		

Note: Values followed by the different letters in the same column are significantly different in the Duncan test  $\alpha$ =0.05; nsnon significant; WAP: week after planting.

Table 4. Effect chicken manure application on antioxidant activity, flavonoid concentration, and flavonoid production in *Pluchea indica*.

Chicken manure	Total flavonoid (mg QUE g <sup>-1</sup> fresh weight)			Antioxidant activity (%) Position			Flavonoid production <sup>2)</sup> (mg QUE per plant)
(kg per plant)							
	3 <sup>rd</sup>	5 <sup>th 1)</sup>	7 <sup>th 1)</sup>	3 <sup>rd</sup>	5 <sup>th</sup>	7 <sup>th</sup>	_
0	1.32	1.07	1.15	58.23	50.07	41.82	29.55
2.5	1.73	1.17	0.93	49.35	37.09	41.15	66.29
5.0	0.92	0.35	0.64	48.41	46.90	44.39	51.45
7.5	1.55	0.94	0.80	47.22	35.89	33.20	71.22
P value	0.100	0.274	0.527	0.147	0.196	0.383	0.382
Sig.	ns	ns	ns	ns	ns	ns	ns

Note: Values followed by the different letters in the same column are significantly different in the Duncan test  $\alpha$ =0.05; ns-non-significant; <sup>1)</sup> Data was transformed using the formula  $\sqrt{x+0.5}$ ; <sup>2)</sup>Data was transformed twice using formula  $\sqrt{x+0.5}$ ; QUE: Quercetin equivalent

nutrient movement (Khan et al., 2023). Potassium in involved is enzyme activation of protein, starch, and adenosine triphosphate (ATP) (Sardans and Peñuelas, 2021). Chicken manure applied to okra resulted in taller plants, more leaves, higher harvest index and fresh weight per plant (Arifah et al., 2019). Other research showed that nitrogen in chicken manure

plays a role in increasing *Vernonia amygdalina* leaf production as indicated by an increase in plant height, number of branches, fresh leaf weight, and dry leaf weight (Tjhia et al., 2018).

Chicken manure applied to alfisol significantly improved the soil nutrient status as shown by

increasing C-organic, available P, exchangeable cations, and the effective cation exchange capacity. In addition, it caused increasing in N, P, and K concentrations in plant tissue, plant height and dry matter yield of soybeans (Soremi et al., 2017). In general, the use of 10 ton.ha-1 of chicken manure has a better effect on most growth variables and yield components, achieving the highest nutrient concentration and nutrient uptake for most macro and micronutrients in onion leaves and bulbs (Falodun and Egharevba, 2018). Similar results were shown when applying 10 tons.ha-1 of chicken manure to Vernonia amygdalina Del. produced a vigor plant and had the highest fresh and dry leaf weights (Ndukwe et al., 2022). Long-term manure application is important for a sustainable environment increasing soil productivity and increasing crop yields (Du et al., 2020).

Provisions of chicken manures can increase soil C-organic and improve soil properties influencing plant growth through increasing soil nutrient cycles so that used as an alternative source of nutrients to increase crop yields, soil fertility, and plant nutrient uptake (Lin et al., 2016). Chicken manure increased the yield and nutrient composition of sweet basil (Yaldız et al., 2019), sorghum (Aziz et al., 2020), and strawberry leaves (Sayğı, 2022). Chicken manure improves tomato performance and nutrient status and increases leaf N, P, K, Ca, and Mg levels (Adekiya and Agbede, 2009). Providing chicken manure made a significant difference to N levels in the 7th leaf position associated with the mobile N element in the tissue. The mature leaves close to the shoot play a role in providing photosynthate for growing shoots or juvenile leaves (Taiz and Zeiger, 2002). The highest nutritional concentration is found in the middle position of cassava leaves (Chaiareekitwat et al., 2022).

Leaf position affected the accumulation of secondary metabolites; phenolics, proanthocyanidins, flavonoids in Lantana camara (Bhakta and Ganjewala, 2009). Younger leaves (positions 1st to 3rd) were most active in the biosynthesis and accumulation of secondary metabolites compared to mature leaves (position 4<sup>th</sup> to 5<sup>th</sup>) (Bhakta and Ganjewala, 2009). Extracts of younger leaf had stronger antioxidant activity than older or mature leaves. Different stages of leaf development had differences in secondary metabolites observed at different leaf positions. The 2<sup>nd</sup> leaf of *Coleus atropurpureus* L. in the vegetative phase had a positive correlation with pigment and total flavonoids (Respita et al., 2019). In general, the total flavonoid concentration in the 3rd leaf position had greater value than the 5th and 7th leaf positions in all treatments as well the antioxidant activity in the 3rd leaf position was supported by the statement (Vongsak et al., 2018) that the leaf shoots of Pluchea indica had

a stronger activity and a higher number of bioactive components compared to mature leaves. Antioxidant properties of P. indica leaves were stronger than Curcuma longa rhizomes (Chan et al., 2022) and green tea Camellia sinensis (Sirichaiwetchakoon et al., 2020). The study showed the total flavonoid concentration at each observed leaf position after the application of chicken manure were similar, and the flavonoid production per plant also showed a similar trend. However, the fresh weight per plant significantly increased by applying chicken manure so in general, the average value of flavonoid production per plant increased with the application of chicken manure. Providing chicken manure no longer showed effects when applied at 45-57 months after planting or after 4-6 harvests. However, heavy pruning at plant height of 60 cm stimulated growth, leaf production, and total flavonoid content of orange jessamine (Jati, et al., 2019). The best level of chicken manure to increase flavonoid production from Vernonia amygdalina was 7.5 kg per plant and increased total flavonoids production by 37% compared to without fertilization (Betty, 2018).

# Conclusion

Application of 5 kg per plant of chicken manures significantly increased *P. indica* fresh leaf weight and dry weight over the lower dose of 2.5 kg per plant or without manures but had no effects on the flavonoid production.

## Acknowledgment

The authors thanked The Indonesian Ministry of Education, Culture, Research and Technology for funding the research entitled "Production of Leaves and Total Flavonoids of *P. indica* (*Pluchea indica* Less.) with Chicken Manure Application" from research scheme BIMA the Master Thesis Research competitive grant in 2023.

## References

Adekiya, A.O., Agbede, T.M. (2009). Growth and yield of tomato (*Lycopersicon esculentum* Mill) as influenced by poultry manure and NPK fertilizer. *Emirates Journal of Food and Agriculture* **21**,10–20. http://cfa.uaeu.ac.ae/ejfa.shtml

Andarwulan, N., Batari, R., Sandrasari, D.A., Bolling, B., and Wijaya, H. (2010). Flavonoid concentration and antioxidant activity of vegetables from

- Indonesia. *Food Chemistry* **121**, 1231–1235. doi:10.1016/j.foodchem.2010.01.033
- Arifah, S.H., Astininngrum, M., and Susilowati, Y.E. (2019). Efektivitas macam pupuk kandang dan jarak tanaman pada hasil tanaman okra (*Abelmaschus esculentus*, L. Moench). *VIGOR* **4**, 38–42. doi:10.31002/vigor.v4i1.1312
- Aziz, A., Khan, B.A., Tahir, M.A., Nadeem, M.A., Amin, M.M., Qura-Tul-Ain, Adnan, M., Munawar, N., Hussain, A., Khisham, M., Toor, M.D., and Sultan, M. (2020). Effect of chicken manure on growth and yield of forage sorghum (Sorghum bicolor L.). International Journal of Botany Studies 5, 401–406. https://www.researchgate.net/publication/342231552
- Bassi, D., Menossi, M., and Mattiello, L. (2018). Nitrogen supply influences photosynthesis establishment along the sugarcane leaf. *Scientific Reports* **8**, 1–13. doi:10.1038/s41598-018-20653-1.
- [Balitttanah] Balai Penelitian Tanah. (2005). "Petunjuk Teknis Analisis Kimia Tanah, Tanaman, Air dan Pupuk". http://palittanah.litbang.deptan.go.id [November 15, 2023].
- Betty. (2018). "Produksi Flavonoids Daun Afrika (*Vernonia amygdalina* Del.) pada Dosis Pupuk Kandang Ayam dan Interval Panen yang Berbeda." [Thesis]. Bogor Agricultural University. Indonesia
- Bhakta, D., and Ganjewala, D. (2009). Effect of leaf positions on total phenolics, flavonoids and proanthocyanidins concentration and antioxidant activities in *Lantana camara* (L). *Journal of Scientific Research* **1**, 363–369. doi:10.3329/jsr.v1i2.1873
- Chaiareekitwat, S., Latif, S., Mahayothee, B., Khuwijitjaru, P., Nagle, M., Amawan, S., and Müller, J. (2022). Protein composition, chlorophyll, carotenoids, and cyanide concentration of cassava leaves (*Manihot esculenta* Crantz) as influenced by cultivar, plant age, and leaf position. *Food Chemistry* 372, 1–9. doi:10.1016/j.foodchem.2021.131173
- Chan, E.W.C., Ng, Y.K., Wong, S.K., and Chan, H.T. (2022). *Pluchea indica*: an updated review of its botany, uses, bioactive compounds and pharmacological properties. *Pharmaceutical Sciences Asia* **49**, 77–85. doi:10.29090/PSA.2022.01.21.113

- Chang, C.C., Yang, M.H., Wen, H.M., and Chern, J.C. (2002). Estimation of total flavonoid concentration in propolis by two complementary colometric methods. *Journal of Food and Drug Analysis* **10**, 178–182. doi:10.38212/2224-6614.2748
- Deng, B., Li, Y., Xu, D., Ye, Q., Liu, G. (2019). Nitrogen availability alters flavonoid accumulation in Cyclocarya paliurus via the effects on the internal carbon/nitrogen balance. *Scientific Reports* **9**, 1–9. doi:10.1038/s41598-019-38837-8.
- Du, Y., Cui, B., Zhang, Q., Wang, Z., Sun, J., and Niu, W. (2020). Effects of manure fertilizer on crop yield and soil properties in China: a meta-analysis. *Catena* **193**, 1–10. doi:10.1016/j. catena.2020.104617
- Falodun, E.J., and Egharevba, R. (2018). Influence of chicken manure rates and spacing on growth, yield, nutrient concentration, uptake and proximate composition of onion (*Allium cepa* L.). *Notulae Scienetia Biologicae* **10**, 117–123. doi:10.15835/nsb10110230
- Fukumoto, L.R., and Mazza, G. (2000). Assessing antioxidant and prooxidant activities of phenolic compounds. *Journal of Agricultural and Food Chemistry* **48**, 3597–3604. doi:10.1021/jf000220w.
- Jati, G.E.A.S., Aziz, S.A., and Melati, M. (2019). Ketinggian pangkas berat dan pupuk organik terhadap biomassa dan flavonoid daun kemuning pada panen pertama dan kedua. *Jurnal Hortikultura Indonesia* **10**,135-144. doi. org/10.29244/jhi.10.2.135-144
- Karimuna, S.R. (2015). "Uji Korelasi Konsentrasi Hara N, P, dan K Daun dengan Produksi Senyawa Bioaktif Kemuning (*Murraya paniculata* (I.) Jack) Akibat Aplikasi Pupuk Kandang Ayam". [Thesis]. IPB University. Indonesia.
- Karimuna, S.R., Aziz, S.A., and Melati, M. (2015). Correlations between leaf nutrient concentration and the production of metabolites in orange jessamine (*Murraya paniculata* L. Jack) fertilized with chicken manure. *Journal of Tropical Crop Science* **2**,16–25. doi:10.29244/jtcs.2.1.16-25.
- [Kementan] Kementerian Pertanian. (2013). "Peraturan Menteri Pertanian Nomor 64/ Permentan/OT.140/5/2013". https://peraturan.

- bpk.go.id/Home/Details/1 60356/permentan -no-64per [February 2, 2023].
- Khan, F., Siddique, A.B., Shabala, S., Zhou, M., and Zhao, C., (2023). Phosphorus plays key roles in regulating plants' physiological responses to abiotic stresses. *Plants* **12**, 1-19. https://doi.org/10.3390/plants12152861
- Komala, O., Andini,S., and Zahra, F. (2020). Uji aktivitas antibakteri sabun wajah ekstrak daun beluntas (*Pluchea indica* L . ) terhadap *Propionibacterium acnes. Fitofarmaka Jurnal Ilmiah Farmasi* **10**,12–21. doi:10.33751/jf.v10i1.1717
- Komala, O., Andini, S., and Zahra, F. (2021). Formulasi dan uji antibakteri sediaan gel serum ekstrak etanol daun beluntas (*Pluchea indica* L.) terhadap Propionibacterium Acne. *Jurnal Buana Farma* 1, 11–13. doi:10.33751/jf.v10i1.1717
- Lestari, K.A.P., Pranoto, P.P., Sofiyah, Musyirah, M., and Pratiwi, F.I. (2020). Antibacterial activity of beluntas (*Pluchea indica* L.) leaves extract using different extraction methods. *Jurnal Riset Biologi dan Aplikasinya* **2**, 49–54. doi:10.26740/jrba.v2n2.p49-54
- Lin, Y., Van Santen, E., and Watts, D. (2016). The effect of chicken litter application on agricultural production: a meta-analysis of crop yield, nutrient uptake and soil fertility *In* "Conference on Applied Statistics in Agriculture". New Prairie Press. Kansas. https://newprairiepress. org/ agstatconference/2016/proceedings/12 [November 18, 2023].
- Marschner, P. (2012). "Marschner's Mineral Nutrition of Higher Plants". Academic Press, 3<sup>rd</sup> edition. San Diego
- Mutrikah, Santoso, H., and Sauqi, A. (2018). Profil bioaktif pada tanaman temulawak (*Curcuma xanthorriza* Roxb) dan beluntas (*Pluchea indica* Less). *Biosaintropis* **4**, 15–21. http://biosaintropis.unisma.ac.id/index.php/biosaintropis/article/view/143/145
- Ndukwe, O.O., Nwakanma, V.T., Orji, C.M., and Iheaturu, D.E. (2022). Chicken manure rate for pot-plant bitter leaf (*Vernonia amygdalina* Del.) production in a humid rainforest zone of Nigeria. *International Journal of Agriculture, Food and Biodiversity* **1**, 25–30.

- Ohtsuki, T., Yokosawa, E., Koyano, T., Preeparame, S., Kowithayakorn, T., Sakai, S., Toida, T., and Ishibashi, M. (2008). Quinic acid esters from Pluchea indica with collagenase, MMP-2 and MMP-9 inhibitory activities. *Phytotherapy Research* **22**, 544–549. doi:10.1002/ptr
- Parker, C. (2012). Pluchea indica (Indian camphorweed). https://doi.org/10.1079/cabicompendium.116400 [January 20, 2024]
- Pranata, N., Boli, G.E.D., Sinta, R., and Sugiaman, V.K. (2021). Effect of beluntas leaf extract (*Pluchea indica*) on oral mucosal wound healing in terms of density of inflammatory cells and collagen. *Systematic Reviews in Pharmacy* **12**, 618–622. doi:10.31838/srp.2021.1.89
- Rasmani. (2021). "Produksi Flavonoid Daun Kelor (*Moringa oleifera* Lam) pada Jarak Tanam dan Jenis Pupuk Kandang yang Berbeda." [Thesis]. Institut Pertanian Bogor. Indonesia.
- Roy, A., Khan, A., Ahmad, I., Alghamdi, S., Rajab, B.S., Babalghith, A.O., Alshahrani, M.Y., Islam, S., and Islam, M.R., (2022). Flavonoids a bioactive compound from medicinal plants and its therapeutic applications. *BioMed Research International* **2022**, 1-8. doi:10.1155/2022/5445291.
- Respita, I.A., Aziz, S.A., and Kurniawati, A. (2019). Correlation of leaf NPK and leaf pigments of Coleus atropurpureus L. Benth during vegetative and generative phases. *Journal of Tropical Crop Science* **6**, 174-181. https://doi.org/10.29244/jtcs.6.03.174-181.
- Sardans, J., and Peñuelas, J. (2021). Potassium control of plant functions: ecological and agricultural implications. *Plants* **10**, 2-31. doi: 10.3390/plants10020419.
- Sayğı, H. (2022). Effects of organic fertilizer application on strawberry (*Fragaria vesca* L.) cultivation. *Agronomy* **12**, 1–15. doi:10.3390/agronomy12051233
- Sirichaiwetchakoon, K., Lowe, G.M., and Eumkeb, G. (2020). The free radical scavenging and anti-isolated human LDL oxidation activities of *Pluchea indica* (L.) Less. tea compared to green tea (*Camellia sinensis*). *BioMed Research International* **2020**, 1-12. doi:10.1155/2020/4183643

- Soremi, A.O., Adetunji, M.T., Adejuyigbe, C.O., Bodunde, J.G., and Azeez, J.O. (2017). Effects of poultry manure on some soil chemical properties and nutrient bioavailability to soybean. *Journal of Agriculture and Ecology Research International* **11**,1–10. doi:10.9734/jaeri/2017/32419.
- Srimoon, R., and Ngiewthaisong, S. (2015). Antioxidant and antibacterial activities of Indian marsh fleabane (*Pluchea indica* (L.) Less). *KKU Research Journal* **20**, 144–154. doi:10.14456/kkurj.2015.12.
- Suriyaphan, O. (2014). Nutrition, health benefits and applications of *Pluchea indica* (L.) less leaves. *Mahidol University Journal of Pharmaceutical Sciences* **41**, 1–10.
- Taiz, L., and Zeiger, E. (2002). "Plant Physiology". Sinauer Associates, Inc, 3<sup>rd</sup> edition. Massachusetts.
- Tariq, H., Asif, S., Andleeb, A., Hano, C., and Abbasi, B.H. (2023). Flavonoid production: current trends in plant metabolic engineering and de novo microbial production. *Metabolites* 13, 1–26. doi:10.3390/metabo13010124.
- Tian, T., Chu, X.Y., Yang, Y., Zhang, X., Liu, Y.M., Gao, J., Ma, B.G., and Zhang, H.Y. (2019). Phosphates as energy sources to expand metabolic networks. *Life* **9**, 2-12. doi: 10.3390/life9020043.
- Tinrat, S. (2021). Phytochemical screening, antioxidant and antimicrobial assessment of *Pluchea Indica* (L.) Less extract as an active ingredient in natural lotion bar. *International Journal of Current Pharmaceutical Research* **13**, (2), 51–57. doi:10.22159/ijcpr.2021v13i2.41555.
- Tjhia, B., Aziz, S.A., and Suketi, K. (2018). Correlations between leaf nitrogen, phosporus and potassium and leaf chlorophyll, anthocyanins and carotenoids content at vegetative and generative stage of bitter leaf (*Vernonia amygealina* Del.) *Journal of Tropical Crop Science* **5**, 25-33. 10.29244/itcs.5.1.25-33

- Vongsak, B., Kongkiatpaiboon, S., Jaisamut, S., and Konsap, K. (2018). Comparison of active constituents, antioxidant capacity, and α -glucosidase inhibition *Pluchea indica* leaf extracts at different maturity stages. *Food Bioscience* **25**, 68–73. doi:10.1016/j. fbio.2018.08.006
- Vongsak, B., Sithisarn, P., Mangmool, S., Thongpraditchote, S., Wongkrajang, Y., and Gritsanapan, W. (2013). Maximizing total phenolics, total flavonoids concentrations and antioxidant activity of Moringa oleifera leaf extract by the appropriate extraction method. *Industrial Crops and Products* **44**, 566–571. doi:10.1016/j.indcrop.2012.09.021
- Walida, H., Harahap, D.E., and Zuhirsyan, M. (2020). Pemberian pupuk kotoran ayam dalam upaya rehabilitasi tanah ultisol Desa Janji yang terdegradasi. *Jurnal Agrica Ekstensia* **14**, 75–80. doi:doi: https://doi.org/10.55127/ae.v14i1.37
- Yaldız, G., Çamlıca, M., Özen, F., and Eratalar, S.A. (2019). Effect of chicken manure on yield and nutrient composition of sweet basil (*Ocimum basilicum* L.). *Communications in Soil Science and Plant Analysis* **50**, 1–16. doi:10.1080/0010 3624.2019.1589488
- Yuliani, Soemarno, Yanuwiadi, B., and Leksono, A.S. (2015). Total phenolic and flavonoid concentrations of *Pluchea indica* less leaves extracts from some altitude habitats. *International Journal of ChemTech Research* 8, 1618–1625.